Synthesis and structure—activity analysis of 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]pyridazin-3(2H)-ones as ligands for benzodiazepine receptors

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Summary — A series of 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]pyridazin-3(2H)-ones were synthesized and evaluated *in vitro* for their affinity toward benzodiazepine receptors (BZRs) in rats and for their intrinsic efficacy in the augmentation of the γ -aminobutyric acid (GABA)-induced chloride currents in the dissociated frog sensory neurons. Compounds in which the 9-position of the condensed-ring system was substituted with alkyl group or bromine had a high affinity toward BZRs. The substituents at the same position also influenced significantly the GABA-induced chloride currents. As the result, 9-alkyl and 9-bromo substituents would interact with the lipophilic area of BZRs. A series of 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]pyridazin-3(2H)-ones exhibited partial and full agonistic activities toward BZRs.

benzodiazepine receptor / y-aminobutyric acid / partial agonist / full agonist / structure-activity relationship

Introduction

Since the discovery of benzodiazepine receptors (BZRs) in the central nervous system [1, 2], the mechanism of the action of benzodiazepine has been rapidly clarified. BZRs act as an allosteric regulatory site of y-aminobutyric acid (GABA) receptors and are considered to mediate two opposite effects: one to amplify or facilitate the action of GABA and the other to reduce it. In respect of these effects, ligands for BZRs have been classified into at least four categories: full agonists, partial agonists, antagonists, and inverse agonists [3, 4]. Most of the drugs in clinical use, such as diazepam 1, are full agonist-type ligands at BZRs and exhibit anticonvulsant, sedative, and muscle relaxant effects in addition to anxiolytic properties. By contrast, the BZR partial agonists were reported as candidates that could maintain the therapeutic potential with fewer side effects than BZR full agonists in the treatment of anxiety [5]. For several years, we have been involved in the design and synthesis of BZR ligands containing a condensed-ring system of pyridazinone as a common molecular element [6–10]. The structure–activity analysis of synthesized compounds verified pharmacologically favorable characteristics of 2-(4-chlorophenyl)-5,6dihydro[1]benzothiepino[5,4-c]pyridazin-3(2H)-one 7-oxide (Y-23684, **2b**) as a BZR partial agonist, namely, an anxioselective anxiolytic agent (fig 1) [9, 11]. Thus, **2b** is now under clinical study in the treatment of anxiety. This finding then led us to an isosteric replacement of the condensed-benzene ring of **2b** with thiophene ring. We herein report the synthesis and evaluation of receptor binding profile of 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]-pyridazin-3(2H)-ones **A** as ligands for BZRs.

Chemistry

We chose a synthetic route to compounds A (fig 1) that focused on the introduction of various substituents

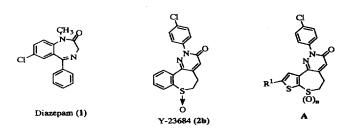


Fig 1. Chemical structures of diazepam **1**, Y-23684 and 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]-pyridazin-3(2*H*)-ones **A**.

into the 9-position of the condensed-ring system and on the oxidation level of sulfide function at the 7-position of the same system. Four kinds of compounds (12a, 13a, 14a and 15a), in which the R₁-substituent at the 9-position of A (n = 0) were hydrogen, methyl, ethyl and propyl groups, respectively, were derived from 6,7-dihydro-5*H*-thieno[2,3-*b*]thiepin-4-ones (5iiv) [12] in a similar manner to in a previous paper of ours [9] (scheme 1). The ketones 5i-iv were prepared by intramolecular cyclization of 4-(2-thienythio)butyric acids (4i-iv), which were obtainable from 2-(lithiomercapto)thiophenes (3i-iv). Compounds 6i-iv were prepared from 5i-iv by the Mannich reaction, and treatment of 6i-iv with excess ammonia, followed by iodomethane to afford corresponding quaternary ammonium salts (7i-iv). Cyanation of 7i-iv and consequent hydrolysis of the products (8i-iv) gave carboxylic acids (9i-iv). These carboxylic acids were then cyclocondensed with 4-chlorophenylhydrazine to produce 2-(4-chlorophenyl)-4,4a,5,6-tetrahydrothieno[2',3':2,3]- thiepino[4,5-c]pyridazin-3(2H)-ones (11i-iv) via intermediate hydrazones (10i-iv). Dehydrogenation [13, 14] with bromine was inappropriate for thiophene derivatives because of the lability of these aromatic rings to electrophiles. We therefore applied a combined treatment [15] with dimethylsulfoxide and hydrogen bromide to the oxidative elimination of hydrogen at the 4- and 4a-positions of 11i, 11ii, 11iii and 11iv, and obtained 12a, 13a, 14a and 15a, respectively.

Some compounds analogous to 12a were synthesized by introducing appropriate substituents into the 9-position of 12a (scheme 2). Thus, 9-bromo compound 17a was prepared by bromination, 9-acyl compounds 18a, 22a, and 23a by Friedel-Crafts reaction, and 9-nitro compound 19a by nitration of 12a. 9-Butyl compound 16a was produced by reduction of 23a with triethylsilane. 9-Formyl compound 21a was obtained by Vilsmeier-Haack reaction of 12a, and 9-cyano compound 20a was prepared by treatment of 21a with hydroxylamine and by subsequent dehydra-

Scheme 1. Synthetic route to 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]pyridazin-3(2H)-ones.

Scheme 2. Modification of the 9-position.

tion of the oxime with trifluoroacetic anhydride. Sulfoxide 12b–20b and sulfone 12c–20c were prepared by oxidation of the corresponding sulfides (scheme 3). Oxidation of the sulfide moiety in the compounds belong to the a series produced corresponding sulfoxide and sulfone derivatives, categorized as b (eg, compound 12b) and c (eg, compound 12c). The 10-propyl derivative of Y-23684 (25b) was prepared from 2a [9] by Friedel–Crafts acylation, reduction of ketone, followed by oxidation of the sulfur atom (scheme 4).

Pharmacological results and discussion

2-(4-Chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-ones were evaluated for

their affinity toward BZRs by an assay on their ability to displace [3H]diazepam from the cerebral cortex of rats [2]. Here, we indicate the BZR affinity of these compounds as their K_i value (nM) in their competition with [3H]diazepam at the binding site. The efficacy of these compounds on BZRs was evaluated in the dissociated frog dorsal root ganglion neurons by use of a concentration-jump (termed 'concentration-clamp') technique, under single-electrode voltage-clamp conditions [16]. Such experiments have demonstrated that all the full agonists for BZRs increase the peak amplitude of chloride currents (I_{Cl}) induced by GABA, and that partial agonists dose-dependently augment this GABA response with an amplitude significantly smaller than diazepam, a typical full agonist [16]. The relative $I_{\rm Cl}$ value (r- $I_{\rm Cl}$), which is the ratio of the $I_{\rm Cl}$ induced by GABA in the presence of the test compound to that induced by GABA itself, has been used for the BZR ligands as a means of predicting the type of activity observed in whole animal models. The chemical structures of synthesized compounds and their in vitro pharmacological data are summarized in table I.

Scheme 3. Oxidation of sulfur atom.

Scheme 4. Synthesis of 10-propyl derivative of Y-23684 (25b).

Table I. Physicochemical and biological data of 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]pyridazin-3(2H)-ones.



Compound	R/a	n	Yield⁰ (%)	Мр (°С)	Recrystallization solvent ^c	Formula	[³H]Diazepam ^d K _i (nM)	Relative $^{ m d}$
12a	Н	0	82	140–142	AcOEt-IPE	C ₁₆ H ₁₁ ClN ₂ OS ₂	6.8 ± 1.0	2.20
12b	Н	1	70	184-186/dec	EtOH-CHCl ₃	$C_{16}H_{11}CIN_2O_2S_2$	54 ± 4	1.34
12c	Н	2	82	277-279	EtOH-CHCl ₃	$C_{16}H_{11}ClN_2O_3S_2$	11 ± 1	1.90
13a	Me	0	62	150-151	EtOH-CHCl ₃	$C_{17}H_{13}ClN_2OS_2$	3.2 ± 0.4	3.64
13b	Me	1	96	174-176/dec	EtOH-CHCl ₃	$C_{17}H_{13}ClN_2O_2S_2$	9.7 ± 0.3	NT^e
13c	Me	2	78	241-243	EtOH-CHCl ₃	$C_{17}H_{13}ClN_2O_3S_2$	2.4 ± 0.7	3.35
14a	Et	0	80	127-128	EtOH-CHCl ₃	$C_{18}H_{15}CIN_2OS_2$	1.9 ± 0.3	3.63
14b	Et	1	87	173-174/dec	EtOH-IPE	$C_{18}H_{15}CIN_2O_2S_2$	1.3 ± 0.3	2.18
14c	Et	2	81	229-231	EtOH-CHCl ₃	$C_{18}H_{15}CIN_2O_3S_2$	1.1 ± 0.7	3.69
15a	Pr	0	82	136-137	EtOH-CHCl ₃	$C_{19}H_{17}CIN_2OS_2$	3.6 ± 1.2	4.37
15b	Pr	1	77	149-151	EtOH-CHCl ₃	$C_{19}H_{17}CIN_2O_2S_2$	1.7 ± 0.4	2.99
15c	Pr	2	65	222-224	EtOH-CHCl ₃	$C_{19}H_{17}CIN_2O_3S_2$	0.84 ± 0.20	4.52
16a	Bu	0	62	125-130	EtOH	$C_{20}H_{19}CIN_2OS_2$	19 ± 5	NTe
16b	Bu	1	59	124-125	EtOH-IPE	$C_{20}H_{19}ClN_2O_2S_2$	8.7 ± 0.3	2.58
16c	Bu	2	83	184–186	EtOH-IPE	$C_{20}H_{19}ClN_2O_3S_2$	4.8 ± 1.1	2.25
17a	Br	0	77	148-151	EtOH-CHCl ₃	$C_{16}H_{10}BrClN_2OS_2$	2.1 ± 0.7	3.92
17b	Br	1	72	184-185/dec	EtOH-CHCl ₃	$C_{16}H_{10}BrClN_2O_2S_2$	0.98 ± 0.23	1.61
17c	Br	2	89	235-237	EtOH-CHCl ₃	$C_{16}H_{10}BrClN_2O_3S_2$	0.61 ± 0.17	1.96
18a	Ac	0	80	251-253	EtOH-CHCl ₃	$C_{18}H_{13}CIN_2O_2S_2$	7.8 ± 1.1	3.94
18b	Ac	1	70	189/dec	EtOH-CHCl ₃	$C_{18}H_{13}CIN_2O_3S_2$	54 ± 5	1.61
18c	Ac	2	80	272-273/dec	EtOH-CHCl ₃	$C_{18}H_{13}CIN_2O_4S_2$	9.9 ± 2.2	2.25
19a	NO_2	0	40	181-182	EtOH-CHCl ₃	$C_{16}H_{10}CIN_3O_3S_2$	11 ± 2	2.46
19b	NO_2	l	78	174-175/dec	EtOH-CHCl ₃	$C_{16}H_{10}CIN_3O_4S_2$	17 ± 2	1.68
19c	NO_2	2	71	280-281/dec	EtOH-CHCl ₃	$C_{16}H_{10}CIN_3O_5S_2$	18 ± 2	1.56
20a	CN	0	60	128-129/dec	EtOH-CHCl ₃	$C_{17}H_{10}CIN_3OS_2$	2.9 ± 0.5	3.66
20 b	CN	1	78	197-199/dec	EtOH-CHCl ₃	$C_{17}H_{10}ClN_3O_2S_2$	14 ± 1	1.39
20c	CN	2	87	292-293	EtOH-CHCl ₃	$C_{17}H_{10}CIN_3O_3S_2$	5.3 ± 0.9	2.12
21a	CHO	0	81	182-183	EtOH-CHCl ₃	$C_{17}H_{11}CIN_2O_2S_2$	1.9 ± 0.1	2.24
22a	PhCO	0	85	180–182	AcOEt	$C_{23}H_{15}ClN_2O_2S_2$	610 ± 20	NTe
1							5.3 ± 1.0	2.54
2b (Y-23684	.)						42 ± 2	1.64

^aPr, *n*-propyl; Bu, *n*-butyl; ^bisolated yield; ^cAcOEt, ethyl acetate; IPE, isopropyl ether; ^dvalues represent the average of three or more experiments. See *Experimental protocols* for details. ^eNT, not tested.

Compound 12b is a simple isostere of 2b at the condensed-ring system, where the benzene moiety was merely replaced with a thiophene ring. These two compounds exhibited a similar BZR affinity. The affinity of 12b was enhanced by reduction or oxidation of its sulfoxide function. The K_i value was 6.8 nM for 12a, 54 nM for 12b, and 11 nM for 12c, but such an order was affected by the existence of the R1-substituent in A. The BZR affinity of A was independent of their partial structure, such as sulfide, sulfoxide and sulfone in the condensed thiophene ring, but was dependent on their structural and the electrostatic properties of the R1-substituent. In comparison with 12a-c, the affinity was commonly enhanced in a series of compounds where the R¹ was an alkyl group or bromine, but other compounds, where the R1 was an electronegative function such as acyl, nitro and cyano, exhibited low binding affinity compared with alkyl compounds. The reduction of binding affinity in benzoyl compound 22a may be attributed to the steric effects caused by the bulky phenyl substituent.

The 9-unsubstituted compounds (12a-c) exhibited smaller r- I_{Cl} values compared with diazepam, as well as 2b. Introduction of alkyl substituents at the 9-position (13a-c-16a-c) remarkably increased the r- I_{Cl} value. Notably, 9-propyl compounds (15a and 15c) possessed an $r-I_{Cl}$ value of more than four. In the case of sulfide compounds, the 9-bromo, 9-acetyl and 9-cyano compounds (17a, 18a and 20a) also exhibited large $r-I_{CI}$ values. On the other hand, introduction of nitro or formyl substituent at the 9-position (19a and 21a) did not give much alteration of the $r-I_{CI}$ value compared with 12a. Moreover, in the case of 9-bromo, 9-acetyl, 9-nitro and 9-cyano compounds, the sulfoxide and sulfone compounds showed smaller r- I_{Cl} values than the sulfide compounds (17a vs 17b and 17c; 18a vs 18b and 18c; 19a vs 19b and 19c; 20a vs 20b and 20c). These results indicated that the 9-substituents would interact with BZRs and affect the intrinsic efficacy of compounds A, and that the oxidation level of the sulfur atom also affected the intrinsic efficacy.

As mentioned above, the 9-substituent was confirmed to be a very important factor in determining not only the affinity of compounds A toward BZRs but also their intrinsic efficacy. This finding prompted us to prepare the 10-propyl derivative of Y-23684 (25b). Our interest was to check whether 10-alkyl substituent in 5,6-dihydro[1]benzothiepino[5,4-c]pyridazin-3(2H)-ones could cause the same alteration. The binding affinity of 25b ($K_i = 7.0 \text{ nM}$) was six times higher than that of 2b, but four times lower than that of 15b. An augmentation of the r- I_{CI} value (the values of 2b and 25b were 1.64 and 2.16, respectively) was smaller than the case of 12b and 15b. Thus, in 5,6-dihydro[1]benzothiepino[5,4-c]pyridazin-3(2H)ones, an influence of introduction of alkyl substituent

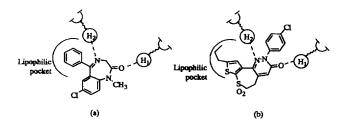


Fig 2. Binding interactions of diazepam 1 (a), 9-propylcompound **15c** (b) with the pharmacophore model [22, 23] for the BZR agonist site. H₁, H₂ are the hydrogen bond donor sites on the receptor protein.

at the 10-position on binding affinity and the r- I_{Cl} value was smaller than the case of thieno compounds A. These results suggest that there is a hydrophobic interaction between alkyl substituent at this position and BZRs, and that π -excessive thiophene ring is necessary to possess high affinity and high intrinsic efficacy.

Based on the studies of SAR and molecular geometry of various ligands, several pharmacophore models [17–23] have been proposed, both agonists and inverse agonists/antagonists. Cook JM et al [22, 23] have reported that BZR agonists interact with two hydrogen bond donating groups termed H₁ and H₂ and with a lipophilic region whose occupation leads to a full agonist. Full occupation of the lipophilic region by phenyl ring (C-5) of diazepam (fig 2a) resulted in a full agonist, while partial occupation of this same region resulted in a partial agonist [24]. In the case of 9-propyl compound 15c, which was classified as a full agonist, this interaction could be mediated by the imino nitrogen atom at the 1-position and carbonyl oxygen atom at 3-position, forming hydrogen bonds with two donor sites (fig 2b). Furthermore, 15c can occupy the proposed lipophilic area with fused thiophene ring and 9-propyl substituent (fig 2b). The partial agonistic properties of 9-bromo, 9-acetyl, 9nitro, 9-cyano and 9-formyl compounds (17b,c-20b,c and **21a**) can be explained by partial occupation of the lipophilic area, or by a negative interaction with the lipophilic area because of their electronegative characters. The oxidation level of sulfur atom should affect the interaction between 9-substituents and BZRs.

To confirm their *in vitro* pharmacological properties we chose representative compounds (14c and 17c) for the *in vivo* tests. Compounds 14c and 17c have the sulfone function as a common structural unit, but are distinguishable only at the R¹-moiety (14c: R¹ = ethyl, 17c: R¹ = Br). These compounds exhibited significantly higher BZR affinity than diazepam. In spite of such a large terminal difference, 14c showed a larger r- I_{C1} value than diazepam and was classified as a full

agonist, but 17c with a smaller r- I_{CI} value was classified as a partial agonist. Such a classification was confirmed in vitro by assessing pharmacological properties in vivo as follows: the ability to prevent bicuculline-induced convulsion in mice (anti-BCL test) [25], the anxiolytic activity determined by the water-lick conflict paradigm in rats (anticonflict test) [26], and the effect on motor coordination in rats (rotarod test). Diazepam 1 was used as a reference drug in these experiments. The pharmacological data are summarized in table II. Compounds 14c, 17c and 1 exhibited a high level of potency in both the anti-BCL and anticonflict tests. In the rotarod test, 14c and 1 were active at doses comparable to the minimum effective doses in the anticonflict test, whereas 17c was inactive at a dose up to 300 mg/kg. These results successfully reflect the in vitro classification of 14c and 17c.

In conclusion, we confirmed that replacement of the fused benzene ring in 2b with a thiophene ring caused a significant change in the affinity and intrinsic efficacy. Introduction of an alkyl or bromo substituent at the 9-position produced a high affinity toward BZRs, and introduction of an electronegative group at the same position caused the r- I_{CI} value to be smaller. This result indicates that the 9-alkyl and 9-bromo substituents would interact with the lipophilic area of BZRs, and that the electronegative 9-substituents would cause a negative interaction. The full occupation of the lipophilic area would lead to a full agonist, whereas a partial occupation or a negative interaction with the lipophilic area would result in a partial agonist. Thus, 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino [4,5-c] pyridazin-3(2H)-ones have a wide spectrum of pharmacological activities, compared with 2-aryl-5,6-dihydro[1]benzothiepino[5,4-c]pyridazin-3(2H)-ones [9] which exhibited partial agonistic properties. 2-(4-Chlorophenyl)-5,6-dihydrothieno-[2',3':2,3]thiepino[4,5-c]pyridazin-3(2H)-ones serve as a tool for exploring the effects of structural modifications on compounds which bind to BZRs, and they should lead to agents useful in the treatment of anxiety and sleep disorders. Studies to evaluate further structural modifications of 2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]pyridazin-3(2H)-one nucleus are currently underway and the results will be published in due course.

Experimental protocols

Chemistry

All melting points were determined on Büchi 530 melting point apparatus, and are uncorrected. Proton nuclear magnetic resonance (¹H-NMR) spectra were recorded with Jeol JNM-EX 270 spectrometer (Me₄Si as an internal standard). Signal multiplicities are represented by s (singlet), d (doublet), t (triplet), q (quartet), brs (broad singlet), and m (multiplet). Chemical shifts are expressed in ppm and coupling constants in hertz

Table II. Biological activity of example compounds.

Compound	Antibicuculline ED ₅₀ (mg/kg, mice, po)	activity MED	Rotarod ED ₅₀ (mg/kg, rats, po)	
14c	0.2	2.5	1.7	
17c	1.4	25	> 300	
1	0.4	10	8.8	

(Hz). The IR spectra were recorded with a Jeol JIR-6500W spectrophotometer. The mass spectra were taken on Jeol JMS-DX 300 system. The elemental analyses were performed for C, H, N, and results were within ±0.4% of the theoretical values. Silica-gel plates (Merck F254) and silica gel 60 (Merck, 70-230 mesh) were used for analytical and column chromatography, respectively.

4-Oxo-4,5,6,7-tetrahydrothieno[2,3-b]thiepin-5-acetic acids **9i-iv** A typical example is given to represent the general procedure.

4-(2-Thienylthio)butyric acid 4i [27]. Under nitrogen, n-butyllithium (1.6 M in hexane, 250 ml, 0.4 mol) was added dropwise to a solution of thiophene (32 g, 0.38 mol) in tetrahydrofuran (500 ml) at -20°C. The solution was stirred at -20°C for an additional 0.5 h, and powdered sulfur (12.8 g, 0.4 mol) was added portionwise below -20°C. The solution was stirred for 0.5 h, and ethyl 4-bromobutyrate (77.9 g, 0.4 mol) was added to the stirred mixture. The solution was kept at 0°C for 1 h and then was brought to room temperature and stirred overnight. Water was added; the layers were separated. The aqueous layer was extracted with ethyl acetate, and the combined ethyl acetate layer and extracts were washed with brine, dried over MgSO₄, and evaporated. The residue was dissolved in EtOH (250 ml). A solution of KOH (30 g, 0.54 mol) in water (250 ml) was added, the mixture stirred for 2 h at room temperature. The solution was made acid and extracted with chloroform, and washed with brine, and concentrated in vacuo. The residue was chromatographed on silica-gel column to give 4i (51 g, 66%); H-NMR (CDCl₃) δ : 1.92 (2H, tt, J = 7.3, 7.3 Hz, SCH₂CH₂CH₂), 2.52 (2H, t, J = 7.3 Hz), 2.84 (2H, t, J = 7.3 Hz) 7.3 Hz), 6.97 (1H, dd, J = 4.0, 5.3 Hz, ArH), 7.12 (1H, d, J =4.0 Hz, ArH), 7.34 (1H, d, J = 5.3 Hz, ArH); IR (KBr) cm⁻¹: 1705 (C=O); MS m/z 202 (M⁺); anal C₈H₁₀O₂S₂ (C, H, N).

6,7-Dihydro-5H-thieno[2,3-b]thiepin-4-one 5i. To a solution of 4i (130 g, 0.64 mol) in toluene (2.0 l) was added with stirring celite (250 g) and phosphorus pentoxide (182 g, 1.28 mol). The mixture was refluxed for 2 h and then filtered. The filtrate was washed with 2% NaHCO₃, dried over MgSO₄, and concentrated *in vacuo* to give 5i (70 g, 59%). Recrystalization from hexane/isopropylether gave colorless crystals, mp 55–57°C (reference 12, mp 53–54°C); 1 H-NMR (CDCl₃) δ : 2.26 (2H, tt, J = 6.6, 6.6 Hz, SCH₂CH₂CH₂), 3.02 (2H, t, J = 6.6 Hz), 3.06 (2H, t, J = 6.6 Hz), 7.08 (1H, d, J = 5.9 Hz, ArH), 7.42 (1H, d, J = 5.9 Hz, ArH); IR (KBr) cm⁻¹: 1655 (C=O); MS m/z 184 (M⁺); anal $C_8H_8OS_2$ (C, H, N).

5-Trimethylammoniomethyl-6,7-dihydro-5H-thieno[2,3-b]thie-pin-4-one iodide 7i. A solution of dimethylamine hydrochloride (69.7 g, 0.85 mol) in 37% HCHO (69.3 g, 0.85 mol)

Table III. Physicochemical data for 4-oxo-4,5,6,7-tetrahydrothieno[2,3-b]thiepin-5-acetic acids.

Compound	$R^{/a}$	Yield⁰ (%)	<i>Mp</i> (° <i>C</i>)	Recrystallization solvent	Formula
9i	Н	18	151–152	EtOH/H ₂ O	$C_{10}H_{10}O_3S_2$
9ii	Me	25	183-186	EtOH/H ₂ O	$C_{11}H_{12}O_3S_2$
9iii	Et	28	180-182	EtOH/H ₂ O	$C_{12}H_{14}O_3S_2$
9iv	Pr	31	177-179	CHCl ₃	$C_{13}H_{16}O_3S_2$

^aPr, *n*-propyl; ^byield from corresponding thiophenes.

Table IV. Physicochemical data for 2-(4-chlorophenyl)-4,4a,5,6-tetrahydrothieno[2',3':2,3]thiepino[4,5-c]pyridazin-3(2H)-ones.

$R^{\prime a}$	Yield⁵ (%)	<i>Mp</i> (° <i>C</i>)	Recrystallization solvent	Formula
Н	83	134–135	EtOH/CHCl ₃	$C_{16}H_{13}CIN_2OS_2$
Me	81	153-154	MeOH/CHCl ₃	$C_{17}H_{15}CIN_2OS_2$
Et	75	101-104	EtOH/CHCl ₃	$C_{18}H_{17}CIN_2OS_2$
Pr	80	Oil		$C_{19}H_{19}ClN_2OS_2$
	H Me Eı	H 83 Me 81 Et 75	H 83 134–135 Me 81 153–154 Et 75 101–104	solvent H 83 134–135 EtOH/CHCl ₃ Me 81 153–154 MeOH/CHCl ₃ Et 75 101–104 EtOH/CHCl ₃

^aPr, *n*-propyl; ^bisolated yield.

was stirred at room temperature for 0.5 h. Acetic anhydride (270 ml) was added dropwise at 70-90°C. After being stirred for 0.5 h, 5i (105 g, 0.57 mol) was added to the mixture at 70°C. The mixture was stirred at 70-75 °C for 3 h. After cooling, the reaction mixture was concentrated in vacuo. The residue was dissolved in chilled water, neutralized with 28% NH₄OH and extracted by CHCl₃. The extract was washed with water, dried over MgSO₄, and evaporated below 40°C. The residue was dissolved in acetone (500 ml), and iodomethane (133 g, 0.93 mol) was added to the resulting solution below 5°C in an ice bath. The mixture was allowed to stand at room temperature for 3 h, and then the crystals formed were collected by filtration and washed with acetone to give 7i (145 g, 66%). Recrystallization from EtOH/H₂O gave colorless crystals, mp 190–192°C; ¹H-NMR (CDCl₃) δ; 1.94–2.06 (1H, m), 2.41– 2.54 (1H, m), 2.68-2.82 (1H, m), 3.06 (9H, s), 3.42-3.67 (2H, m), 3.80-3.90 (1H, m), 4.28-4.38 (1H, m), 7.36 (1H, d, J =5.3 Hz, ArH), 7.45 (1H, d, J = 5.3 Hz, ArH); IR (KBr) cm⁻¹: 1670 (C=O); anal C₁₂H₁₈INOS₂ (C, H, N).

4-Oxo-4,5,6,7-tetrahydrothieno[2.3-b]thiepin-5-acetonitrile 8i. To a solution of 7i (145 g, 0.38 mol) in methanol (500 ml) was added a solution of KCN (60 g, 0.92 mol) in water (150 ml) dropwise at room temperature. The solution was stirred at room

temperature for 1 h and poured into ice-water. The resulting mixture was extracted with CHCl₃. The extract was washed with water, dried over MgSO₄, and concentrated *in vacuo*. After the addition of isopropyl ether to the residue, the crystals formed were collected by filtration (75 g, 89%). Recrystallization from EtOH gave **8i** as colorless crystals, mp 68–70°C; ¹H-NMR (CDCl₃) δ : 1.99–2.11 (1H, m), 2.53–2.90 (4H, m), 3.31–3.40 (1H, m), 3.71–3.76 (1H, m), 7.12 (1H, d, J = 5.3 Hz, ArH), 7.46 (1H, d, J = 5.3 Hz, ArH); IR (KBr) cm⁻¹: 2250 (CN), 1655 (C=O); MS m/z 223 (M⁺); anal $C_{10}H_{9}NOS_{2}$ (C, H, N).

4-Oxo-4,5.6,7-tetrahydrothieno[2,3-h]thiepin-5-acetic acid **9i**. To a solution of conc HCl (300 ml) and acetic acid (300 ml) was added **8i** (75 g, 0.34 mol). The solution was refluxed for 4 h, and poured into ice-water. The precipitate was collected by filtration, washed with water, and recrystallized from EtOH/H₂O to give **9i** (63.8 g, 78%) as colorless needles, mp 151–152°C; ¹H-NMR (CDCl₃) &: 1.92–2.05 (1H, m), 2.27–2.42 (1H, m), 2.53 (1H, dd, J = 17.2, 4.6 Hz), 2.73–2.85 (1H, m), 3.08 (1H, dd, J = 17.2, 8.6 Hz), 3.24–3.33 (1H, m), 3.75–3.86 (1H, m), 7.08 (1H, d, J = 5.3 Hz, ArH), 7.41 (1H, d, J = 5.3 Hz, ArH), 8.70 (1H, brs, COOH); IR (KBr) cm⁻¹: 1705

(COOH), 1660 (C=O); MS m/z 242 (M+); anal $C_{10}H_{10}O_3S_2$ (C, H, N).

The other compounds (9ii-iv) in table III were prepared in a similar manner.

2-(4-Chlorophenyl)-4,4a,5,6-tetrahydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-ones 11i-iv
A typical example is given to represent the general procedure.

2-(4-Chlorophenyl)-4,4a,5,6-tetrahydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-one IIi. A mixture of 9i (58.7 g, 0.24 mol), sodium acetate (24 g, 0.29 mol), and 4-chlorophenyl-hydrazine halfsulfate (55.8 g, 0.29 mol) in EtOH (800 ml) was refluxed for 15 h. After evaporation of the solvent, the residue was dissolved in acetic acid (800 ml). The mixture was refluxed for 3 h, poured into ice-water, and extracted with CHCl₃. The extract was washed with water, dried over MgSO₄, and concentrated in vacuo. The residual solid was recrystalized with EtOH/CHCl₃ give 11i (69.5 g, 83%) as a pale yellow powder, mp 134–135°C; ¹H-NMR (CDCl₃) δ : 2.10–2.19 (1H, m), 2.25–2.32 (1H, m), 2.64 (1H, dd, J = 16.5, 2.0 Hz), 2.92–3.06 (2H, m), 3.12–3.21 (1H, m), 3.76–3.83 (1H, m), 7.13 (1H, d, J = 5.3 Hz, H-10), 7.33 (1H, d, J = 5.3 Hz, H-9), 7.36 (2H, d, J = 8.6 Hz, ArH), 7.55 (2H, d, J = 8.6 Hz, ArH); IR (KBt) cm⁻¹: 1695 (C=O); MS m/z: 348 (M⁺); anal $C_{16}H_{13}ClN_2OS_2$ (C, H, N).

The other compounds (11ii–iv) in table IV were prepared in a similar manner.

2-(4-Chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]-pyridazin-3(2H)-ones 12a-15a

A typical example is given to represent the general procedure.

2-(4-Chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]-pyridazin-3(2H)-one 12a. To a solution of 11i (69 g, 0.20 mol) in acetic acid containing 15% HBr (400 ml) was added dropwise dimethylsulfoxide (14.8 ml, 0.21 mol) at room temperature. The reaction mixture was stirred for 0.5 h, and poured into ice-water, and extracted with CHCl₃. The extract was washed with water and 2% NaHCO₃, dried over MgSO₄, and concentrated in vacuo. The residual solid was recrystallized with ethyl acetate/isopropyl ether to give 12a (56 g, 82%) as a pale yellow powder, mp 140–142°C; ¹H-NMR (CDCl₃) δ t. 2.85 (2H, t, J = 6.6 Hz, SCH₂CH₂), 3.34 (2H, t, J = 6.6 Hz, SCH₂CH₂), 6.95 (1H, s, C=CHCO), 7.30 (1H, d, J = 5.3 Hz, H-10), 7.33 (1H, d, J = 5.3 Hz, H-9), 7.44 (2H, d, J = 8.6 Hz, ArH), 7.67 (2H, d, J = 8.6 Hz, ArH); IR (KBr) cm⁻¹: 1680 (C=O); MS m/z: 346 (M⁺); anal C₁₆H₁₁ClN₂OS₂ (C, H, N).

The other compounds (13a-15a) in table I were prepared in a similar manner.

2-(4-Chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4.5-c|-pyridazin-3(2H)-one 7-oxides 12b-20b

A typical example is given to represent the general procedure.

2-(4-Chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4.5-c]-pyridazin-3(2H)-one 7-oxide 12b. To a stirred solution of 12a (0.55 g, 1.6 mmol) in acetic acid (20 ml) was added dropwise 30% H_2O_2 (0.5 g, 4.4 mmol) below 10°C. The mixture was stirred at room temperature for 6 h, poured into ice-water, and extracted with CHCl₃. The extract was washed with 2% NaHSO₃ and water, dried over MgSO₄, and concentrated in vacuo. The residual solid was recrystallized with EtOH/CHCl₃ to give 12b (0.4 g, 70%) as a colorless powder, mp 184–186°C (dec); ¹H-NMR (CDCl₃) δ : 2.98 (1H, ddd, J = 14.5, 7.9, 4.6 Hz, SCH₂CH₂). 3.36 (1H, ddd, J = 14.5, 7.9, 4.0 Hz,

SCH₂CH₂), 3.50 (1H, ddd, J = 13.9, 7.9, 4.0 Hz, SCH₂CH₂), 3.67 (1H, ddd, J = 13.9, 7.9, 4.6 Hz, SCH₂CH₂), 6.98 (1H, s, C=CHCO), 7.44 (2H, d, J = 8.6 Hz, ArH), 7.56 (1H, d, J = 5.3 Hz, H-10), 7.64 (2H, d, J = 8.6 Hz, ArH), 7.68 (1H, d, J = 5.3 Hz, H-9); IR (KBr) cm⁻¹: 1675 (C=O); MS m/z: 362 (M⁺); anal C₁₆H₁₁ClN₂O₂S₂ (C, H, N).

The other compounds (13b-20b) in table I were prepared in a similar manner.

2-(4-Chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]-pyridazin-3(2H)-one 7,7-dioxides 12c-20c
A typical example is given to represent the general procedure.

2-(4-Chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thiepino[4,5-c]-pyridazin-3(2H)-one 7,7-dioxide 12c. To a stirred solution of 12a (1.0 g, 2.9 mmol) in CH_2Cl_2 (20 ml) was added 80% m-chloroperbenzoic acid (1.3 g, 6.0 mmol) below 10°C. The mixture was stirred at room temperature for 5 h. The resulting solution was washed with 2% NaHCO₃, dried over MgSO₄, and concentrated in vacuo. The residual solid was recrystallized with EtOH/CHCl₃ to give 12c (0.9 g, 82%) as colorless needles, mp 277–279°C; ¹H-NMR (CDCl₃) &: 3.04 (2H, t, J = 6.6 Hz, SCH₂CH₂), 7.24 (1H, s, C=CHCO), 7.55 (1H, d, J = 5.3 Hz, H-10), 7.58 (2H, d, J = 8.6 Hz, ArH), 7.72 (2H, d, J = 8.6 Hz, ArH), 8.15 (1H, d, J = 5.3 Hz, H-9); IR (KBr) cm⁻¹: 1675 (C=O); MS m/z: 378 (M+); anal $C_{16}H_{11}ClN_2O_3S_2$ (C, H, N).

The other compounds (13c-20c) in table I were prepared in a similar manner.

9-Bromo-2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-one **17a**

To a stirred solution of 12a (2.0 g, 5.8 mmol) in acetic acid (80 ml) was added bromine (1.0 g, 6.3 mmol) at 10°C. The reaction mixture was kept at room temperature for 2 h, and was then poured into ice-water. The precipitate was collected by filtration, washed with H_2O , recrystallized from EtOH/CHCl₃ to give 17a (1.9 g, 77%) as colorless needles, mp 148–151°C; ¹H-NMR (CDCl₃) δ : 2.87 (2H, t, J = 6.6 Hz, SCH₂CH₂), 3.35 (2H, t, J = 6.6 Hz, SCH₂CH₂), 6.94 (1H, s, C=CHCO), 7.27 (1H, s, H-10), 7.44 (2H, d, J = 9.2 Hz, ArH), 7.65 (2H, d, J = 9.2 Hz, ArH); IR (KBr) cm⁻¹: 1680 (C=O); MS m/z: 426 (M+); anal $C_{16}H_{10}BrClN_2OS_2$ (C, H, N).

9-Acyl-2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-ones 18a, 22a, 23a A typical example is given to represent the general procedure.

9-Butyryl-2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-one 23a. To an ice-cooled suspension of AlCl₃ (3.8 g, 29 mmol) in CH₂Cl₂ (50 ml) was added butyrylchloride (0.9 g, 8.4 mmol) and the mixture was stirred at 0–10°C for 0.5 h. After addition of 12a (2.0 g, 5.8 mmol), the mixture was refluxed for 1 h and then poured into ice-water. The resulting mixture was extracted with CH₂Cl₂. The extract was washed with water, dried over MgSO₄. and concentrated in vacuo. The residual solid was recrystallized with EtOH/CHCl₃ to give 23a (1.5 g, 62%) as colorless needles, mp 152–153°C; ¹H-NMR (CDCl₃) δ : 0.99 (3H, t, J = 7.3 Hz, CH₃CH₂CH₂CO), 2.84 (2H, tt, J = 7.3, 7.3 Hz, CH₃CH₂CH₂CO), 2.93 (2H, t, J = 6.6 Hz, SCH₂CH₂), 3.42 (2H, t, J = 6.6 Hz, SCH₂CH₂), 6.96 (1H, s, C=CHCO), 7.46 (2H, d, J = 8.6 Hz, ArH), 7.66 (2H, d, J = 8.6 Hz, ArH), 7.83 (1H, s, H-10); IR (KBr) cm⁻¹: 1650 (C=O), 1670 (C=O); MS m/z: 388 (M+); anal C₂₀H₁₇ClN₂O₂S₂ (C, H, N).

The other compounds (18a and 22a) were prepared in a similar manner.

9-Butyl-2-(4-chlorophenyl)-5,6-dihydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-one 16a

To a stirred solution of **23a** (1.0 g, 2.4 mmol) in trifluoroacetic acid (15 ml) was added triethylsilane (0.6 g, 5.2 mmol) at room temperature. The reaction mixture was stirred at room temperature for 15 h, poured into ice-water, and extracted with CHCl₃. The extract was washed with 2% NaHCO₃, dried over MgSO₄, and concentrated *in vacuo*. The residue was chromatographed on a silica gel using CHCl₃ as an eluent to give **16a** (0.6 g, 62%), which was recrystallized from EtOH to afford a colorless powder, mp 125–130°C; ¹H-NMR (CDCl₃) δ : 0.94 (3H, t, J = 7.3 Hz, CH₃CH₂CH₂CH₂), 1.40 (2H, tq, J = 7.3, 7.3 Hz, CH₃CH₂CH₂CH₂), 2.78 (2H, t, J = 7.3, CH₃CH₂CH₂CH₂), 2.78 (2H, t, J = 7.3, CH₃CH₂CH₂CH₂), 2.84 (2H, t, J = 6.6 Hz, SCH₂CH₂), 6.93 (1H, s, C=CHCO), 6.98 (1H, s, H-10), 7.44 (2H, d, J = 8.6 Hz, ArH), 7.66 (2H, d, J = 8.6 Hz, ArH); IR (KBr) cm⁻¹: 1680 (C=O); MS m/z: 402 (M+); anal C₂₀H₁₉CIN₂OS₂ (C, H, N).

2-(4-Chlorophenyl)-9-nitro-5,6-dihydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-one **19a**

To a stirred solution of 12a (2.5 g, 7.2 mmol) in acetic acid (100 ml) and acetic anhydride (1 ml) was added fuming HNO₃ (0.6 ml) at 10°C. The reaction mixture was stirred for 0.5 h, followed by poured into ice-water. The resulting mixture was extracted with CHCl₃. The extract was washed with water, dried over MgSO₄, and concentrated *in vacuo*. The residue was chromatographed on a silica gel using CHCl₃ as an eluent to give 19a (1.3 g, 40%), which was recrystallized from EtOH/CHCl₃ to afford a yellow powder, mp $181-182^{\circ}$ C; ¹H-NMR (CDCl₃) δ : 2.98 (2H, t, J = 6.6 Hz, SCH₂CH₂), 3.48 (2H, t, J = 6.6 Hz, SCH₂CH₂), 6.97 (1H, s, C=CHCO), 7.45 (2H, d, J = 8.6 Hz, ArH), 7.64 (2H, d, J = 8.6 Hz, ArH), 8.09 (1H, s, H-10); IR (KBr) cm⁻¹: 1670 (C=O); MS m/z: 391 (M+); anal $C_{16}H_{10}$ ClN₃O₃S₂ (C, H, N).

2-(4-Chlorophenyl)-9-formyl-5,6-dihydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-one 21a

A solution of phosphorus oxychloride (3.5 g, 22.8 mmol) and *N*-methylformanilide (3.1 g, 22.8 mmol) was stirred for 0.5 h at room temperature. To the solution was added **12a** (4.0 g, 11.5 mmol), the mixture was stirred for 8 h at room temperature and was then poured into water. The resulting mixture was extracted with CHCl₃. The extract was washed with water, dried over MgSO₄, and concentrated *in vacuo*. The residue was chromatographed on silica gel using CHCl₃ as an eluent to give **21a** (3.5 g, 81%), which was recrystallized from EtOH/CHCl₃ to afford a colorless powder, mp 182–183°C; 1 H-NMR (CDCl₃) &: 2.97 (2H, t, J = 6.6 Hz, SCH₂CH₂), 3.45 (2H, t, J = 6.6 Hz, SCH₂CH₂), 6.96 (1H, s, C=CHCO), 7.45 (2H, d, J = 8.6 Hz, ArH), 7.65 (2H, d, J = 8.6 Hz, ArH). 7.95 (1H, s, H-10), 9.82 (1H, s, CHO). IR (KBr) cm⁻¹: 1665 (C=O); MS m/z: 374 (M+); anal $C_{17}H_{11}$ ClN₂O₂S₂ (C, H, N).

2-(4-Chlorophenyl)-9-cyano-5,6-dihydrothieno[2',3':2,3]thie-pino[4,5-c]pyridazin-3(2H)-one **20a**

To a stirred solution of **21a** (1.0 g, 2.7 mmol) in EtOH (50 ml) was added hydroxylamine hydrochloride (0.3 g, 4.3 mmol) and NaHCO₃ (0.36 g, 4.3 mmol) at room temperature. The reaction mixture was refluxed for 5 h and was then poured into icewater. The resulting mixture was extracted with CHCl₃. The extract was washed with water, dried over MgSO₄, and concen-

trated *in vacuo*. The residue was dissolved in tetrahydrofuran (10 ml), and triethylamine (0.8 g, 7.9 mmol) and trifluoroacetic anhydride (0.56 g, 2.7 mmol) were added to the resulting solution below 5°C in an ice bath. The reaction mixture was stirred for 1 h, and was then poured into ice-water. The resulting mixture was extracted with CHCl₃. The extract was washed with water, dried over MgSO₄, and concentrated *in vacuo*. The residual solid was recrystallized with EtOH/CHCl₃ to give **20a** (0.6 g, 60%) as colorless needles, mp 128–129°C (dec); 'H-NMR (CDCl₃) δ : 2.92 (2H, t, J = 6.6 Hz, SCH₂CH₂), 3.44 (2H, t, J = 6.6 Hz, SCH₂CH₂), 6.98 (1H, s, C=CHCO), 7.45 (2H, d, J = 8.6 Hz, ArH), 7.63 (2H, d, J = 8.6 Hz, ArH). 7.79 (1H, s, H-10); IR (KBr) cm⁻¹: 2225 (CN), 1680 (C=O); MS m/z: 371 (M+); anal C₁₇H₁₀ClN₃OS₂ (C, H, N).

2-(4-Chlorophenyl)-10-propionyl-5,6-dihydro[1]benzothie-pino[5,4-c]pyridazin-3(2H)-one **24a**

To an ice-cooled suspension of AlCl₃ (5.9 g, 44 mmol) in 1,2dichloroethane (100 ml) was added propionylchloride (1.2 g, 13 mmol) and the mixture was stirred at 0-10°C for 0.5 h. After addition of 2a (3.0 g, 8.8 mmol), the mixture was stirred for 2 h at 60°C and then poured into ice-water. The resulting mixture was extracted with CHCl3. The extract was washed with water, dried over MgSO₄, and concentrated in vacuo. The residue was chromatographed on a silica gel using CHCl₃ as an eluent to give 24a (1.8 g, 52%), which was recrystallized from EtOH/CHCl₃ to afford colorless needles, mp 197-198°C; ¹H-NMR (CDCl₃) δ : 1.23 (3H, t, J = 7.3 Hz, CH_3CH_2CO), 2.75 $(2H, t, J = 6.6 \text{ Hz}, \text{SCH}_2\text{C}H_2), 3.04 (2H, q, J = 7.3 \text{ Hz},$ $CH_3CH_2CO)$, 3.30 (2H, t, J = 6.6 Hz, SCH_2CH_2), 6.92 (1H, s, C=CHCO), 7.45 (2H, d, J = 9.2 Hz, ArH), 7.68 (2H, d, J =9.2 Hz, ArH), 7.73 (1H, d, J = 7.9 Hz, ArH), 7.98 (1H, dd, J =2.0, 7.9 Hz, ArH), 8.13 (1H, d, J = 2.0 Hz, ArH); IR (KBr) cm⁻¹: 1680 (C=O); MS m/z: 396 (M+); anal $C_{21}H_{17}ClN_2O_2S$ (C, H, N).

2-(4-Chlorophenyl)-10-propyl-5,6-dihydro[1]benzothiepino-[5,4-c]pyridazin-3(2H)-one **25a**

This compound was synthesized from **24a** in the same way as described for the preparation of **16a**, mp 178–179°C; ¹H-NMR (CDCl₃) δ : 0.96 (3H, t, J = 7.3 Hz, CH₃CH₂CH₂), 1.66 (2H, tq, J = 7.3, 7.3 Hz, CH₃CH₂CH₂), 2.63 (2H, t, J = 7.3 Hz, CH₃CH₂CH₂), 2.72 (2H, t, J = 6.6 Hz, SCH₂CH₂), 3.22 (2H, t, J = 6.6 Hz, SCH₂CH₂), 6.89 (1H, s, C=CHCO), 7.23 (1H, t, J = 2.0, 7.9 Hz, ArH), 7.40 (1H, d, J = 2.0 Hz, ArH), 7.44 (2H, d, J = 8.6 Hz, ArH), 7.53 (1H, d, J = 7.9 Hz, ArH), 7.68 (2H, d, J = 8.6 Hz, ArH); IR (KBr) cm⁻¹: 1670 (C=O); MS m/z: 382 (M+); anal C₂₁H₁₉ClN₂OS (C, H, N).

2-(4-Chlorophenyl)-10-propyl-5,6-dihydro[1]benzothiepino-[5,4-c]pyridazin-3(2H)-one 7-oxide **25b**

This compound was synthesized from 25a in the same way as described for the preparation of 12b, mp 201–202°C; ¹H-NMR (CDCl₃) δ : 0.98 (3H, t, J = 7.3 Hz, CH₃CH₂CH₂), 1.70 (2H, tq, J = 7.3, 7.3 Hz, CH₃CH₂CH₂), 2.68–2.90 (4H, m), 3.07–3.17 (1H, m), 3.85–3.97 (1H, m), 6.96 (1H, s, C=CHCO), 7.44 (2H, d, J = 8.6 Hz, ArH), 7.46 (1H, d, J = 2.0 Hz, ArH), 7.53 (1H, d, J = 2.0, 7.9 Hz, ArH), 7.67 (2H, d, J = 8.6 Hz, ArH), 7.82 (1H, d, J = 7.9 Hz, ArH); IR (KBr) cm⁻¹: 1675 (C=O); MS mz: 398 (M+); anal C₂₁H₁₉ClN₂O₂S (C, H, N).

Pharmacology

Benzodiazepine receptor binding assay

Preparation of a synaptosome fraction and [3H]diazepam binding studies were carried out according to the method of

Möhler and Okada [2]. Crude synaptosomal membranes were suspended in a 50 mM Tris-HCl buffer (pH 7.4) containing 120 mM NaCl and 5 mM KCl. The reaction was started by the addition of a 900 µl aliquot of crude synaptosomal membranes to 100 µl solution containing [3H]diazepam (final concentration was 2 nM) and a known concentration of the test compounds. After the mixture was incubated for 20 min at 0°C, the binding was stopped by addition of 3 ml of ice-cold 50 mM Tris-HCl buffer (pH 7.4) containing 120 mM NaCl and 5 mM KCl. The samples were then filtered under vacuum through Whatman GF/B filters and immediately washed 4 times with 3 ml of icecold buffer. The radioactivity on the filters was measured by a liquid scintillation counter Binding in the presence of 1 μ M unlabelled diazepam was defined as non-specific binding. Specific binding was defined as the difference between the total binding and the non-specific binding. The K_i values were determined by the relationship $K_i = IC_{50}/(1 + c/K_d)$, where IC_{50} was the concentration of the test compounds which caused a 50% reduction of the specific binding vs the control, c was the concentration of [^{3}H]diazepam (2 nM), and K_{d} was the dissociation constant determined by Scatchard's plot. The K_i values are means \pm SE of at least three determinations.

GABA-induced chloride current in frog sensory neuron

The experiment was carried out according to the published method [16]. Bull frog dorsal root ganglion neurons were isolated. Isolated neuronal cell bodies were perfused internally and externally by a suction pipette technique with respective test solutions for recording the chloride current. The external solution contained Tris-HCl 89, CsCl 2, MgCl₂ 5, TEA-Cl 25, glucose 5, and HEPES 10 (each unit, mM) and was adjusted at pH 7.4 with appropriate Tris-base. The internal solution contained CsCl 95, Cs-aspartate 10, TEA-Cl 25, HEPES 10 and EGTA 2.5 (each unit, mM) and was adjusted to pH 7.2. Neurons were voltage-clamped at a holding membrane potential of -50 mV with a single electrode. Test compounds were applied by using a concentration-clamp technique. Augmentative action of test compound on the GABA response was examined on $I_{\rm Cl}$ induced by 3×10^{-6} M GABA. The results were presented as relative values of peak $I_{\rm Cl}$ elicited by 3 \times 10^{-6} M GABA alone. The relative I_{Cl} values represented the mean of at least three determinations and the SE for these values were generally $\leq 10\%$ of the mean.

Anticonvulsant test (antibicuculline test)

The experiment was a modification of the method of Lippa and Regan [25]. Groups of 7–14 ddY male mice were challenged with bicuculline (0.6 mg/kg iv) 1 h after the oral administration of the test compounds. The ED₅₀ values were calculated by the probit method as the dose which prevented tonic extension in half of the animals.

Anticonflict test (water-lick test)

The experiment was carried out by a modification of the method of Vogel et al [26]. Groups of 10–14 male Wister rats were deprived of water for 72 h, and were placed in the test chamber and allowed to drink from the water spout. Licking was automatically accompanied by a 100 V, 0.2–0.3 mA,

300 ms electrical stimulus across the grid floor and spout every 20 licks. After the rat received the first electrical stimulus, the number of stimuli was recorded automatically during the subsequent 3 min test. The test compounds were administered orally 1 h before the test. The MED was defined as the lowest dose to produce a statistically significant increase in the punished responses compared with control.

Rotarod test

Groups of 10–14 male Wister rats were used. The rats were gently placed on the rod (5 cm in diameter rotating at 5 rpm) 1h after oral administration of the test compounds. The $\rm ED_{50}$ value was calculated by the probit method as the dose which caused half of the animals to drop from the rotarod within

References

- 1 Squires RF, Braestrup C (1977) Nature (Lond) 266, 732
- 2 Möhler H, Okada T (1977) Life Sci 20, 2101-2110
- 3 Petersen EN (1987) Drugs Future 12, 1043-1053
- 4 Müller WE (1988) Drugs Today 24, 649-663
- 5 Haefely WE, Martin JR, Schoch P (1990) Trends Pharmacol Sci 11, 452-456
- 6 Nakao T, Kawakami M, Morita K, Morimoto Y, Takehara S, Tahara T (1990) Yakugaku Zasshi 110, 561-572
- 7 Nakao T, Kawakami M, Morita K et al (1990) Yakugaku Zasshi 110, 573-585
- 8 Nakao T, Tanaka H, Morimoto Y, Takehara S, Demizu K, Tahara T (1990) Yakugaku Zasshi 110, 922–931
- 9 Nakao T, Obata M, Kawakami M et al (1991) Chem Pharm Bull Jpn 39, 2556-2563
- 10 Nakao T, Obata M, Yamaguchi Y, Marubayashi N, Ikeda K, Morimoto Y (1992) Chem Pharm Bull Jpn 40, 117-121
- 11 Yasumatsu H, Morimoto Y, Yamamoto Y et al (1994) Br J Pharmacol 111, 1170-1178
- 12 Baldwin JJ, Ponticello GS (1986) Jpn Kokai Tokkyo Koho JP 61158978; Chem Abstr 106, 33016v
- 13 Sterk EA, Brundage RP, Fletcher LT (1953) J Am Chem Soc 75, 1117-1119
- 14 Curran WV, Ross A (1974) J Med Chem 17, 273–281
- 15 Nakao T, Obata M, Yamaguchi Y, Tahara T (1991) Chem Pharm Bull Jpn 39, 524–526
- 16 Yakushiji T, Fukuda T, Oyama Y, Akaike N (1989) Br J Pharmacol 98, 735, 740
- 17 Tebib S, Bourguignon JJ, Wermuth CG (1987) J Comput-Aided Mol Des 31, 153-170
- 18 Borea PA, Gilli G, Bertolasi V, Ferretti V (1987) Mol Pharmacol 31, 334-344
- 19 Allen MS, Hagen TJ, Trudell ML, Codding PW, Skolnick P, Cook JM (1988) J Med Chem 31, 1854–1861
- 20 Villar HO, Uyeno ET, Toll L, Polgar W, Davies MF, Loew GH (1989) Mol Pharmacol 36, 586-600
- 21 Villar HO, Davies MF, Loew GH, Maguire PA (1991) Life Sci 48, 593-602
- 22 Hollinshead SP, Trudell ML, Skolnick P, Cook JM (1990) J Med Chem 33, 1062-1069
- 23 Zhang W, Koehler KF, Harris B, Skolnick P, Cook JM (1994) J Med Chem 37, 745–757
- 24 Diaz-Arauzo H, Evoniuk GE, Skolnick P, Cook JM (1991) J Med Chem 34, 1754, 1756
- 25 Lippa AS, Regan B (1977) Life Sci 21, 1779-1784
- 26 Vogel JR, Beer B, Clody DC (1971) Psychopharmacologia 21, 1-12
- 27 Cagniant P, Cagniant D (1966) Bull Soc Chim Fr 2172-2179